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Toward new camptothecins. Part 4: On the reactivity of nitro and amino precursors of aza analogs of 5-methoxycarbonyl camptothecin

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Abstract—In the context of formation of new aza analogs of camptothecin, nitration then reduction of condensed pyridones was realized, leading to new derivatives of pyrrolo-aza-indoles. Treatment of these compounds with hydrobromic acid led to new structure rearrangements while oxidation of the α -position was unsuccessful. Mechanisms of formation of the new products are discussed. © 2007 Elsevier Ltd. All rights reserved.

1. Introduction

The 2-pyridone unit is a key structural feature of a number of biologically active compounds.¹ Some pyridones act as antiviral,² or as herbicides,³ pesticides,⁴ fungicides,⁵ antibiotics,⁶ or antioxidant agents.⁷ On the other hand, it can be found in the toxin ricinine,⁸ the psychoactive huperzine A,⁹ or in the lead anticancer alkaloid camptothecin (Fig. 1).

The isolation and structure of camptothecin were reported in 1966 by Wall^{10a} and many reviews, devoted to this compound, revealed its paramount importance.^{10b,c,12} Interest in this cytotoxic drug and semisynthetic analogs was stimulated when its mode of action was discovered. When the cleavable complex between topoisomerase I and DNA is stabilized by camptothecin, collision of the replication fork with this reversible complex¹¹ leads to cell death by preventing DNA religation.¹² Irinotecan and topotecan have emerged from bioavaibility and tolerance studies, and are used in cancer treatment.¹³ Recently, the crystal structure of the DNA-topoisomerase I-camptothecin complex was solved and two models of camptothecin-DNA-topoisomerase I interaction were formulated.¹⁴ In the camptothecin series, structure-activity relationships are well established in regard to modifications in A, B, C, and D rings. They



Figure 1. Structures of some active pyridones.

also indicated the ability of substituents in positions 7, 9, 10, and 11 to maintain or improve biological activity. The need for a hydroxy lactone ring E has been established as a golden rule¹⁵ and, for instance, lactams **1** (Fig. 2) possess poor activity.¹⁶ However, this dogma fell with the discovery of other compounds stabilizing the complex between DNA and topoisomerase I in the same way as camptothecin: the homocamptothecins such as the diflomotecan, or the cyclopentanones such as S39625 are now in clinical phase.¹⁷ The luotonin is also now known to exhibit one tenth of the camptothecin topo I inhibitory activity (Fig. 2).¹⁸

The targets of the present work were lactams 2 designed to explore the potential of aza analogs of cyclopentanones such as S39625 (Fig. 2). We have already described the synthesis of 5-methoxycarbonyl camptothecin 3 from DL methyl pyroglutamate 4.^{19c} A key intermediate of this reaction scheme was ketone 5, obtained by oxidation of enamine 6 (Scheme 1). Interest in the methoxycarbonyl group of 2 came from its position face to face with the major groove

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Figure 2. Camptothecin analogs.

in the potential complex with DNA and topo I. It can probably be replaced easily by a hydrogen,^{19a} thereby increasing the size of the library of compounds to be tested biologically.



Scheme 1. Synthesis of 5-methoxycarbonyl camptothecin analog.

2. Results and discussion

2.1. Synthesis of nitro compounds and treatment of products in acidic media

As a first approach to the reaction of nitration, pyridone 8^{19a} was submitted to the action of AcONO₂, although the

preferential reactivity of these condensed pyridones toward electrophiles in the *para* position of the carbonyl was known.²⁰ Compound 9 nitrated at the *ortho* position was not observed in the reaction media, and only products 10–12 were isolated by chromatography on SiO₂. The syntheses of mono and dinitro products 10 and 12 were not unexpected, but formation of the *gem* disubstituted compound 11 was interesting, highlighting the particular reactivity at the α -position of condensed pyridones (Scheme 2).

Because mono nitropyridone **9** was not formed during this first reaction, nitration of triester **7** was now realized. Only small amounts of nitrated triester **13** were isolated after using 63% HNO₃/AcOH or HNO₃/H₂SO₄; however, HNO₃/TFAA²¹ led to 79% of **13**, and HNO₃/Ac₂O²² gave quantitative yield of the same product (Scheme 3).

Removal of the aromatic methoxycarbonyl group of **13** was then studied. Such a decarboxylation of esters of condensed pyridones is well known, realized by reflux in 48% HBr. Thereby triester **13** was treated in these conditions (Scheme 3). The remaining ester functions were hydrolyzed in the course of the reaction and the compounds need to be reesterified; that was realized with MeOH, while drying the ternary azeotrope H₂O/MeOH/CHCl₃ on molecular sieves.^{19a} Compounds **14–17** were obtained with a 95% total yield.

2.2. Formation of pyridone 14

Removal of the nitro group of the previous nitropyridone **13** is only a consequence of the reversibility of the reaction of electrophiles with aromatics. An interesting result is the better



Scheme 2. Reaction conditions: (i) (a) 48% HBr, reflux, 5 h; (b) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h;^{19a} (ii) Ac₂O/63% HNO₃, 60 °C, 2 h.



Scheme 3. Reaction conditions: (i) Ac₂O/63% HNO₃, 60 °C, 1 h; (ii) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h.

yield of **14** obtained by using 30% HCl. This led very cleanly to 90% of **14**, while 4% H₂SO₄ gave 70% of the same product, accompanied by low amounts of diester **15** and lactone **16**. These better yields were not caused by the rise in the acidity because 30% HCl is less acidic than 48% HBr. So this increase in **14** was due to the decrease in the formation of the other by-products. This interpretation of the results was validated when KBr was added to HBr, leading to the preferential formation of lactone **16**, to the disadvantage of **14** (Scheme 3).

2.3. Formation of oxidized products 15 and 16

Pyridones **15** and **16** came from an oxidation of the lateral chain of **13**. In a possible mechanism that was due to oxygen

dissolved in hydrobromic acid²³ even when the reactions were performed under nitrogen. Because triester **7** (or the corresponding triacid) was not oxidized in the same conditions¹⁹ (Scheme 4), the nitro group of **13** was necessary for the oxidation to occur. As the presence of KBr increased the yield in lactone **16** (Scheme 3), the Br⁻ anion probably mediated this reaction. Alcohol **18** was thus obtained more easily from triester **13** or triacid **19** (Scheme 4). In this product, the hydroxyacid chain was substituted in the α -position of the acid by the strongly withdrawing nitropyridone group. Decarboxylation of acid **18**, then cyclization of alcohol **20** led to lactone **16**. On the other hand, alcohol **18** can evolve to ketone **21**. Indeed, decarbonylation of α -amino (amido, oxo) acid derivatives **22** to give oxonium ions **23** is



Scheme 4. Reaction conditions: (i) 48% HBr, reflux, 5 h; (ii) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h.^{19a}

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a well-known reaction²⁴ (Scheme 5). When R=H (22), the oxonium formed was the protonated form of ketone 21. Another oxidation step changed the ethyl ketone chain of 21 to a new carboxylic acid 24; protolysis of the nitro group, decarboxylation of the *para*-carboxyacid, and reesterification led then to pyridone 15.



Scheme 5. Oxonium ions.

2.4. Formation of oxidized products 17

A first light on the reaction mechanism leading to compound **17** was obtained in the following reaction: in the hope to clarify the possible influence of ester groups, triester **13** was first heated in sodium hydroxide. The resulting crude product was heated at reflux in 48% HBr then reesterified in the usual way. Only new pyridone **25** was isolated in 30% yield, without traces of phenol **17**. A possible mechanism involves initial nucleophilic attack of the hydroxyl ion on the carbon carrying the nitro function, with formation of the anionic σ^{H} adduct **26** (Meisenheimer complex). Upon reflux in HBr, the pyridone ring of triacid **27** would be open, to give ketone **28** (drawn in its enolic form in Scheme 6). Recyclization, oxidative aromatization, and decarboxylation would then furnish diacid **29**, whose esterification gave **25**.

In this process, the crucial need for the aromatic carboxylic group of **27** to open the pyridone ring was highlighted by the fact that analog **30**, obtained from saponification of diester **17**, does not evolve toward **25** (Scheme 6).

Formation of compound 17 can now be described as the opening of the pyridone ring of triacid 31 then hydrolysis of nitro compound 32 to give ketone 33 (these two compounds are drawn in their enolic forms in Scheme 6). Because 33, unlike 28, did not yield diester 25, the

decarboxylation of **32** occurred before hydrolysis of the nitro group.

2.5. Reduction then decarboxylation of nitropyridone 13

Because decarboxylation of triester 13 does not yield the required nitropyridone, it was decided to perform this step after reduction of the nitro function. Treatment of 13 either with Pd/C or with Zn/HBr yields 83 or 70%, respectively, of azaindolinedione 34, thus furnishing the E ring of compound 2. Decarboxylation of 34 was realized as previously by reflux in 48% HBr then reesterification of acid 36 with MeOH. Ester 36 was not obtained, but methoxyazaindolinone 37 was isolated in 86% yield, accompanied by 2% of amine 38. Iminoether 39, formed from 36 and MeOH/H⁺, could be an intermediate in the formation of 37. On the other hand, oxidation of heterocycle 35 or of the aminoacid obtained from hydrolysis of the lactam ring of 35, probably yielded 40. As shown in Scheme 7, loss of carbon monoxide would then lead to aminoketones 41 then 38. Acid 35 was characterized by NMR, but was not isolated; if needed, it will be necessary to proceed to its esterification by using other methods that the reflux in MeOH/H⁺.

2.6. Attempts to introduce a ketone group in C ring

Many reactions were performed in the aim to introduce a ketone functionality in the pyrrolidine C ring. Some of them are reported in Scheme 8; the use of selenium oxide was earlier reported by Danishefsky²⁰ in his synthesis of camptothecin, and we have already described the oxidation of enedimethylamines in the same context.¹⁹ Only one result needs some comments: heating nitropyridone 13 with Bredereck's reagents easily gave 92% of enamine 42. Oxidation of this compound by NaIO₄ led to entire degradation of the reaction medium. Because that could be caused by oxidative cleavage of the enol form of ketone 43, stabilization of the keto form was attempted by complexation with CeCl₃. The obtained mixture led rapidly to a very complex NMR spectrum (there were three asymmetric centers in the new product obtained) but some peaks could originate from an aldehyde or its enol form. The crude mixture was then



Scheme 6. Reaction conditions: (i) 48% HBr, reflux, 5 h; (ii) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h; (iii) NaOH, reflux 6 h.



Scheme 7. (i) Zn/HBr/MeOH, 20 °C; 1 h; (ii) 48% HBr, reflux, 24 h; (iii) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h.



Scheme 8. α-Oxidation of C ring.

treated by acetic anhydride, giving a quantitative yield of enol acetate 44.

2.7. NMR spectra of new compounds

Many of the new products obtained in this study were unexpected compounds. Elucidation of their structures was



Figure 3. ¹H NMR spectra of pyridones 11, 15–17, 29, and 39 and assignments determined by 2D NMR techniques: COSY, NOESY, HMBC, and HSQC.

carried out by NMR, mass spectrometry (LC–MS), infra-red and elemental analyses. Some of ¹H NMR spectra as well as HMBC and HSQC correlations are reported in Figure 3.

3. Conclusion

Introduction of the ketone functionality in the pyrrolidine C ring of the key precursors of aza analogs of 5-methoxycarbonyl camptothecin was unsuccessful. Thus it will be necessary to use other methods than nitration to obtain these compounds. However, some of the new products described in this study could give dipeptide analog scaffolds useful in the field of rigid β -turn peptidomimetics (Fig. 4).²⁵



Figure 4. Analogy between 34, 38, and 39 and rigid β-turn peptidomimetics.

4. Experimental section

4.1. General

Melting points were determined using an Electrothermal apparatus and are uncorrected. ¹H and ¹³C NMR spectra were obtained on a Varian Gemini 2000 at 200 and 50 MHz, respectively. IR spectra were obtained in ATR mode on a FTIR Bruker Tensor 27. Thin layer chromatographies were performed on precoated Kieselgel 60F₂₅₄ plates. APCI⁺ (atmospheric pressure chemical ionization) mass spectra were obtained on an LC–MS system Thermo Electron Surveyor MSQ. Microanalyses were performed by the 'Service de Microanalyses' of LSEO, Université de Bourgogne, Dijon, France. All products are obtained as mixtures of diastereoisomers.

4.2. Nitration of pyridone 8

Nitric acid (63%, 2 mL) was slowly added to a solution of pyridone **8** (900 mg, 3 mmol) in acetic anhydride (11 mL) at 0 °C. The mixture was stirred at 60 °C for 1 h and the solvent was evaporated. The residue was diluted with EtOAc (50 mL) and washed with satd aqueous NaHCO₃ (50 mL). The aqueous phase was extracted with EtOAc (3×30 mL) and the combined organic layers were dried by MgSO₄ then evaporated. The residue was purified by chromatography on SiO₂ (heptane/EtOAc, 100/0% to 0/100%), to give compounds **10–12**.

4.2.1. Methyl 7-(1-(methoxycarbonyl)propyl)-1,2,3,5-tetrahydro-6,8-dinitro-5-oxoindolizine-3-carboxylate (10). Recrystallized from methanol/ether; yellow powder; 40% yield; TLC R_f (EtOAc)=0.7; mp 65–67 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.92, 0.93 (2t, J=7.5 and 7.5 Hz, 3H, CHCH₂CH₃), 1.52–1.78 (m, 2H, CHCH₂CH₃), 2.20–2.69 (m, 2H, CH₂CH₂CH), 3.44–3.78 (m, 3H, CH₂CH₂CH, CHCH₂CH₃), 3.65 (s, 3H, CO₂CH₃), 5.19, 5.22 (2dd, J=9.6, 3.3 Hz and J=8.0, 1.6 Hz, 1H, CH₂CH₂CH); ¹³C NMR (CDCl₃, 50 MHz) δ 12.2 (CH₃), 25.0 (CH₂), 25.4 (CH₂), 33.4 (CH₂), 48.7 (CH), 52.2 (CH₃), 53.0 (CH₃), 62.3 (CH), 118.2 (C), 146.0 (C), 153.3 (C), 159.2 (C), 169.3 (C), 171.3 (C), 171.6 (C); IR: ν cm⁻¹ 1745, 1680, 1534, 1338. Anal. Calcd for C₁₅H₁₇O₉N₃: C, 47.00; H, 4.47; N, 10.96. Found: C, 46.92; H, 4.66; N, 10.63.

4.2.2. Methyl 7-(1-(methoxycarbonyl)propyl)-1,2,3,5-tetrahydro-1,1,8-trinitro-5-oxoindolizine-3-carboxylate

(11). Recrystallized from EtOAc/ether; yellow powder; 10% yield; mp 105–107 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.81, 0.83 (2t, *J*=7.3 Hz and *J*=7.3 Hz, 3H, CH₂CH₃), 1.71–2.20 (m, 2H, CH₃CH₂CH), 3.65, 3.67 (2s, 3H, CO₂CH₃), 3.68 (t, *J*=7.3 Hz, 1H, CHCH₂CH₃), 3.75, 3.76 (2s, 3H, CO₂CH₃), 3.78–4.14 (m, 2H, CH₂CH), 5.43, 5.45 (2dd, *J*=9.5, 1.7 Hz and *J*=9.4, 1.6 Hz, 1H, CH₂CH₂CH), 7.3, 7.31 (2s, 1H, CH); ¹³C NMR (CDCl₃, 50 MHz) δ 11.6 (CH₃), 24.4 (CH₂), 24.8 (CH₂), 35.5 (CH), 47.9 (CH₃), 53.0 (CH₃), 106.4 (C), 119.4 (C), 139.6 (CH), 139.7 (C), 144.9 (C), 151.8 (C), 167.7 (C), 170.4 (C); IR: ν cm⁻¹ 1741, 1683, 1632, 1543, 1358; LC–MS (APCI⁺) *m*/*z* 429 (MH⁺). Anal. Calcd for C₁₅H₁₆N₄O₁₁: C, 42.06; H, 3.77; N, 13.08. Found: C, 42.38; H, 3.80; N, 13.48.

4.2.3. Methyl 7-(1-(methoxycarbonyl)propyl)-1,2,3,5-tetrahydro-8-nitro-5-oxoindolizine-3-carboxylate (12). Recrystallized from methanol/ether; yellow powder; 35% yield; TLC R_f (EtOAc)=0.8; ¹H NMR (CDCl₃, 200 MHz) δ 0.99, 1 (2t, J=7.5 Hz and J=7.5 Hz, 3H, CH₂CH₃), 1.75– 2.27 (m, 1H, CH₂CH₂CH), 2.3–2.7 (m, 1H, CH₂CH₂CH), 3.55–3.67 (m, 2H, CH₂CH₂CH), 3.71 (s, 3H, CO₂CH₃), 3.83 (s, 3H, CO₂CH₃), 3.92, 3.94 (2t, J=7.1 Hz and J=7.2 Hz, 1H, CHCH₂CH₃), 5.18 (dd, J=9.3, 3.4 Hz, 1H, CH₂CH₂CH), 6.44 (s, 1H, CH); ¹³C NMR (CDCl₃, 50 MHz) δ 11.8 (CH₃), 24.9 (CH₂), 27.3 (CH₂), 29.4 (CH₂), 45.1 (CH), 52.0 (CH₃), 52.8 (CH₃), 61.2 (CH), 100.6 (CH), 126.7 (C), 138.3 (C), 141.7 (C), 156.5 (C), 170.1 (C), 173.6 (C). Anal. Calcd for C₁₅H₁₈O₇N₂: C, 53.25, H, 5.36, N, 8.28. Found: C, 53.11; H, 5.42; N, 8.08.

4.3. Dimethyl 7-[1-(methoxycarbonyl)propyl]-6-nitro-5oxo-1,2,3,5-tetrahydro-3,8-indolizinedicarboxylate (13)

Nitric acid (63%, 90 mL, 0.6 mol) was slowly added to a solution of compound 7 (70 g, 0.2 mol) in acetic anhydride (1 L) at 0 °C. The mixture was stirred at 60 °C for 1 h and the solvent was evaporated. The residue was diluted with EtOAc (50 mL) and washed by satd aqueous NaHCO₃ (50 mL). The aqueous phase was extracted with EtOAc $(3 \times 30 \text{ mL})$ and the combined organic layers were dried (MgSO₄). The residue obtained upon evaporation was crystallized from methanol/ ether; yellow powder; 95% yield; TLC R_f (EtOAc)=0.6; mp 147–149 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.92, 0.95 (2t, J=7.3 Hz and J=7.6 Hz, 3H, CH₂CH₃), 1.5-1.7 (m, 2H, CHCH₂CH₃), 2.25-2.65 (m, 2H, CH₂CHCO), 3.35-3.65 (m, 3H, CHCH₂CH₃, CH₂CH₂CH), 3.68, 3.69 (2s, 3H, CO₂CH₃), 3.79, 3.80 (2s, 3H, CO₂CH₃), 3.83, 3.84 (2s, 3H, CO₂CH₃), 5.2, 5.24 (2dd, J=9.6, 3.5 Hz and J=9.6, 2.9 Hz, 1H, NCHCO₂CH₃); ¹³C NMR (CDCl₃, 50 MHz) δ 12.6 (CH₃), 23.7 (CH₂), 24.1 (CH₂), 25.5 (CH₂), 33.3 (CH₂), 48.1 (CH), 52.1 (CH₃), 52.4 (CH₃), 53.3 (CH₃), 62.6 (CH), 104.6 (C), 144.0 (C), 152.7 (C), 157.3 (C), 164.6 (C), 169.1 (C), 170.4 (C); IR: ν cm⁻¹ 1740, 1723, 1660, 1605, 1533, 1433. Anal. Calcd for C₁₇H₂₀O₉N₂: C, 51.51; H, 5.02; N, 6.79. Found: C, 51.52; H, 5.09; N, 7.07.

4.4. Decarboxylation of nitro compound 13

A stirred solution of pyridone 7 (1 g, 2.5 mmol) in 48% hydrobromic acid (50 mL) was heated at 130 °C for 48 h then evaporated. Methanesulfonic acid (0.10 mg) in methanol (200 mL) and chloroform (200 mL) were added to the

residue, and the mixture was refluxed for 48 h while drying the solvent by condensing it in a Soxhlet-type apparatus containing 3 Å molecular sieves (50 g). Dichloromethane (200 mL) was added to the residue obtained upon evaporation, and the solution was washed with aqueous NaHCO₃. The organic phase was dried (MgSO₄) then evaporated. The residue was purified by chromatography on SiO₂ (heptane/EtOAc, 100/0% to 0/100%) to give pyridones **14–17**.

4.4.1. Methyl 7-[1-(methoxycarbonyl)propyl]-5-oxo-1.2.3.5-tetrahydroindolizine-3-carboxylate (14). Recrystallized from EtOAc; white powder; 48% yield; TLC R_f (EtOAc)=0.45; mp 65–67 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.91, 0.92 (2t, J=7.2 Hz and J=7.6 Hz, 3H, CHCH₂CH₃), 1.7-2.1 (m, 2H, CHCH₂CH₃), 2.2-2.6 (m, 2H, CH₂CH₂CH), 2.9–3.2 (m, 2H, CH_2CH_2CH), 3.28 (t, J=7.7 Hz, 1H, CHCH₂CH₃), 3.69, 3.69 (2s, 3H, CO₂CH₃), 3.8 (s, 3H, CO₂CH₃), 5.08, 5.09 (2dd, J=9.1, 2.9 Hz and J=5.9, 3.2 Hz, 1H, CH₂CH₂CH), 6.15 (s, 1H, ArH), 6.3 (s, 1H, ArH); ¹³C NMR (CDCl₃, 50 MHz) δ 11.8 (CH3), 25.5 (CH₂), 26.1 (CH₂), 30.3 (CH₂), 52.1 (CH), 52.6 (CH3), 52.9 (CH3), 60.9 (CH), 101 (CH), 116.4 (CH), 150 (C), 152.6 (C), 161.3 (C), 170.4 (C), 172.8 (C); IR: ν cm⁻¹ 1727, 1657, 1589, 1532, 1432, 1204; Anal. Calcd for C₁₅H₁₉NO₅: C, 61.49; H, 5.91; N, 5.17. Found: C, 61.49; H, 6.78; N, 5.17.

4.4.2. Dimethyl 1,2,3,5-tetrahydro-5-oxoindolizine-3,7dicarboxylate (**15**). White powder; 23% yield; TLC R_f (EtOAc)=0.4; ¹H NMR (CDCl₃, 200 MHz) δ 2.24–2.74 (m, 2H, CH₂CH₂CH), 3.03–3.36 (m, 2H, CH₂CH₂CH), 3.79 (s, 3H, CO₂CH₃), 3.9 (s, 3H, CO₂CH₃), 5.13 (dd, J=9.4, 3.1 Hz, 1H, CH₂CH₂CH), 6.63, 7.03 (2s, ArH, 2H); ¹³C NMR (CDCl₃, 50 MHz) δ 8.4 (CH₃), 25.8 (CH₂), 27.4 (CH₂), 29.5 (CH₂), 52.9 (CH₃), 61.6 (CH), 81.2 (CH), 102.3 (C), 108.7 (CH), 155.1 (C), 159.3 (C), 161.1 (C), 167.4 (C), 169.6 (C); LC–MS (APCI⁺) m/z 251 (MH⁺). Whatever the purification method, this product was always contaminated with 10% of **14**.

4.4.3. Methyl 3-ethyl-1,3,5,7,8,9-hexahydro-1,5-dioxofuro[3,4-g]indolizine-7-carboxylate (16). Recrystallized from EtOAc/ether; white powder; 23% yield; TLC R_f (EtOAc)=0.4; mp 112–114 °C; ¹H NMR (CDCl₃, 200 MHz) δ 1.03 (t, J=7.4 Hz, 3H, CHCH₂CH₃), 1.69–2.16 (m, 2H, CHCH₂CH₃), 2.36–2.75 (m, 2H, CH₂CH₂CH), 3.34–3.69 (m, 2H, CH₂CH₂CH), 3.82 (s, 3H, CO₂CH₃), 5.17 (dd, J=9.6, 3.2 Hz, 1H, CH₂CH₂CH), 5.31, 5.34 (2dd, J=7.0, 1.4 Hz and J=7.0, 1.4 Hz, 1H, CHCH₂CH₃), 6.29 (m, 1H, ArH); ¹³C NMR (CDCl₃, 50 MHz) δ 8.4 (CH₃), 25.8 (CH₂), 27.4 (CH₂), 29.5 (CH₂), 52.9 (CH₃), 61.6 (CH), 81.2 (CH), 102.3 (C), 108.7 (CH), 155.1 (C), 159.3 (C), 161.1 (C), 167.4 (C), 169.6 (C); IR: ν cm⁻¹ 1742, 1677; LC–MS (APCI⁺) m/z 278 (MH⁺). Anal. Calcd for C₁₄H₁₅O₅N: C, 60.65; H, 5.45; N, 5.05. Found C, 60.60; H, 5.60; N, 5.08.

4.4.4. Methyl 7-(1-(methoxycarbonyl)propyl)-1,2,3,5-tetrahydro-6-hydroxy-5-oxoindolizine-3-carboxylate (17). Recrystallized from EtOAc; white cotton; 1% yield; TLC R_f (EtOAc)=0.55; mp 172–174 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.91, 0.93 (2t, J=7.4 Hz and J=7.4 Hz, 3H, CHCH₂CH₃), 1.59–1.85 (m, 1H, CHCH₂CH₃), 1.9–2.15 (m, 1H, CHCH₂CH₃), 2.23–2.65 (m, 2H, CH₂CH₂CH), 3.68, 3.7 (2s, CO₂CH₃, 3H), 3.8, 3.81 (2s, CO₂CH₃, 3H), 3.93, 3.95 (2t, *J*=7.7 Hz and *J*=7.7 Hz, 1H, C*H*CH₂CH₃), 5.11, 5.13 (2dd, *J*=7.4, 1.8 Hz and *J*=7.4, 1.8 Hz, 1H, CH₂CH₂C*H*), 6.16 (m, 1H, ArH), 6.96 (br s, 1H, OH); ¹³C NMR (CDCl₃, 50 MHz) δ 11.8 (CH₃), 24.9 (CH₂), 27.3 (CH₂), 29.4 (CH₂), 45.1 (CH), 52.0 (CH₃), 52.8 (CH₃), CH (61.2), 100.6 (CH), 126.7 (C), 138.3 (C), 141.7 (C), 156.5 (C), 170.1 (C), 173.6 (C); IR: ν cm⁻¹ 1751, 1734, 1677, 1340, 1211; LC–MS (APCI⁺) *m*/*z* 310 (MH⁺). Anal. Calcd for C₁₅H₁₉O₆N: C, 8.25; H, 6.19; N, 4.53. Found: C, 58.25; H, 6.24; N, 4.74.

4.5. Dimethyl 6-ethyl-5-oxo-1,2,3,5-tetrahydroindolizine-3,7-dicarboxylate (29)

A stirred solution of pyridone 7 (500 mg, 1, 3 mmol) in 30% sodium hydroxide (20 mL) was heated at 70 °C for 16 h. The mixture was acidified by 48% hydrobromic acid (50 mL) and was heated at 130 °C for 48 h then evaporated. Methanesulfonic acid (0.10 mg) in methanol (200 mL) and chloroform (200 mL) were added to the residue, and the mixture was refluxed for 48 h while drying the solvent by condensing it in a Soxhlet-type apparatus containing 3 Å molecular sieves (50 g). Dichloromethane (200 mL) was added to the residue obtained upon evaporation, and the solution was washed with aqueous NaHCO₃. The organic phase was dried (MgSO₄) then evaporated. The residue obtained upon evaporation was crystallized from methanol/ ether; yellow powder; 45% yield; ¹H NMR (CDCl₃, 200 MHz) δ 1.14 (t, J=7.4 Hz, 3H, CH₂CH₃), 2.22-2.61 (m, 2H, CHCH₂CH₂), 2.97-3.30 (m, 2H, CH₂CH₂CH), 3.80 (s, 3H, CO₂CH₃), 3.90 (s, 3H, CO₂CH₃), 5.11 (dd, J=9.3 Hz and J=3.5 Hz, 1H, CH₂CH₂CH₂; ¹³C NMR (CDCl₃, 50 MHz) & 13.4 (CH₃), 21.7 (CH₂), 26.3 (CH₂), 30.3 (CH₂), 52.4 (CH₃), 52.7 (CH₃), 61.6 (CH), 99.9 (CH), 133.9 (C), 138.9 (C), 146.9 (C), 161.4 (C), 167.3 (C), 170.4 (C); LC-MS (APCI+) m/z 280 (MH+). Anal. Calcd for C14H17O5N: C, 60.21; H, 6.14; N, 5.02. Found: C, 59.67; H, 6.21; N, 4.89.

4.6. Methyl 3-ethyl-2,9-dioxo-2,3,5,6,7,9-hexahydro-1*H*-pyrrolo[2,3-*f*]indolizine-4,7-dicarboxylate (34)

Zinc dust (600 mg) was slowly added to a stirring solution of pyridone 7 (400 mg, 1 mmol) in HBr (48%, 25 mL) and methanol (2 mL). After 1 h, the mixture was filtered on Celite and the filtrate was diluted with satd aqueous NaHCO₃ (50 mL). The aqueous layer was extracted by EtOAc $(3 \times 100 \text{ mL})$ and the combined organic layers were dried. Residue obtained upon evaporation was purified by crystallization in methanol. Recrystallized from methanol; white powder; 70% yields; TLC R_f (EtOAc)=0.56; mp 174-176 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.83 (t, J=7.3 Hz, 3H, CH₂CH₃), 1.93-2 (m, 2H, CHCH₂CH₃), 2.3-2.66 (m, 2H, CH₂CHCO), 3.48–3.66 (m, 2H, CH₂CH₂CH), 3.83 (s, CO₂CH₃), 3.85–3.88 (m, 4H, CO₂CH₃ and 3H. CHCH₂CH₃), 5.27 (dd, J=3.3, 9.5 Hz, 1H, NCHCO₂CH₃), 8.6 (s, 1H, NH); ¹³C NMR (CDCl₃, 50 MHz) δ 9.2 (CH₃), 23.4 (CH₂), 26.0 (CH₂), 32.5 (CH₂), 50.7 (CH), 51.7 (CH₃), 53.0 (CH₃), 61.6 (CH), 104.4 (C), 130.5 (C), 135.6 (C), 151.2 (C), 151.4 (C), 164.7 (C), 169.9 (C), 178.6 (C); IR: v cm⁻¹ 3224, 1729, 1717, 1653, 1576, 1537, 1438, 1212. Anal. Calcd for C₁₅H₁₇O₉N₃: C, 47.00, H, 4.47, N, 10.96. Found: C, 46.92; H, 4.66; N, 10.63.

4.7. Decarboxylation of compound 34

A stirred solution of pyridone **34** (6 g, 18 mmol) in 48% hydrobromic acid (120 mL) was heated at 130 °C for 24 h then evaporated. Methanesulfonic acid (0.25 mg) in methanol (200 mL) and chloroform (200 mL) were added to the residue and the solution was refluxed for 48 h while drying the solvent by condensing it in a Soxhlet-type apparatus containing 3 Å molecular sieves (50 g). Dilute NaHCO₃ (200 mL) was added to the residue obtained upon evaporation and the solution was extracted with dichloromethane (2×400 mL). The combined organic layers were dried (MgSO₄) then evaporated. The residue was purified by chromatography on SiO₂ (heptane/EtOAc, 100/0% to 0/100%) to give pyridones **37** and **38**.

4.7.1. Methyl 3-ethyl-5,6,7,9-tetrahydro-2-methoxy-9oxo-1H-pyrrolo[2,3-f]indolizine-7-carboxylate (37). Recrystallized from methanol/ether; gray powder; 97% yield; TLC R_f (EtOAc)=0.15; mp 235–237 °C; ¹H NMR (CDCl₃, 200 MHz) δ 1.16 (t, J=7.5 Hz, 3H, CH₂CH₃), 2.22–2.62 (m, 2H CH₂CH₂CH), 2.52 (q, J=7.5 Hz, 2H, CH₂CH₃), 2.97-3.32 (m, 2H, CH₂CH₂CH), 3.74 (s, 3H, CO₂CH₃), 4.08 (s, 3H, OCH₃), 5.17 (dd, J=9.4, 2.6 Hz, 1H, CH₂CH₂CH), 6.37 (t, J=1.4 Hz, 1H, ArH); ¹³C NMR (CDCl₃, 50 MHz) δ 15 (CH₂), 15.8 (CH₃), 27.4 (CH₂), 29.6 (CH₂), 52.5 (CH₃), 59.3 (CH₃), 60.5 (CH), 95.7 (CH), 99.4 (C), 114.2 (C), 133.5 (C), 139.7 (C), 152.0 (C), 152.5 (C), 171.5 (C); IR: v cm⁻¹ 1750, 1663, 1587, 1567, 1198; LC-MS (APCI⁺) m/z 291 (MH⁺). Anal. Calcd for C₁₅H₁₈O₄-N₂·H₂O: C, 58.43; H, 6.54; N, 9.09. Found: C, 58.25; H, 6.24: N. 9.60.

4.7.2. Methyl 6-amino-1,2,3,5-tetrahydro-5-oxo-7-propionylindolizine-3-carboxylate (38). Yellow foam; 2% yield; TLC R_f (EtOAc)=0.6; ¹H NMR (CDCl₃, 200 MHz) δ 1.19 (t, J=1.4 Hz, 3H, CH₂CH₃), 2.22–2.61 (m, 2H CH₂CH₂CH), 2.52 (q, J=7.4 Hz, 2H, CH₂CH₃), 2.90–3.22 (m, 2H, CH₂CH₂CH), 3.79 (s, 3H, CO₂CH₃), 5.11 (dd, J= 9.0, 3.1 Hz, 1H, CH₂CH₂CH), 6.4 (t, J=1.6 Hz, 1H, ArH), 7.24 (br s, 2H, NH₂); ¹³C NMR (CDCl₃, 50 MHz) δ 8.1 (CH₃), 27.0 (CH₂), 28.8 (CH₂), 32.6 (CH₂), 52.6 (CH₃), 60.9 (CH), 99.4 (CH), 112.7 (C), 131.3 (C), 140.4 (C), 157.2 (C), 170.3 (C), 202.0 (C). Anal. Calcd for C₁₄H₁₇O₄N: C, 59.08; H, 6.10; N, 10.60. Found: C, 59.64; H, 6.42; N, 10.02.

4.8. Dimethyl (1*E*)-7-(1-(methoxycarbonyl)propyl)-1-((dimethylamino)methylene)-1,2,3,5-tetrahydro-6nitro-5-oxoindolizine-3,8-dicarboxylate (42)

A stirred mixture of triester **13** (300 mg, 0.75 mmol) and Bredereck's reagent (210 mg, 1.05 mmol) was heated at 110 °C for 3 h (N₂). The residue obtained upon evaporation was diluted in water (40 mL) then extracted by dichloromethane (2×100 mL). The residue was dissolved in dichloromethane, Celite was added, and the solvent was evaporated. The powder was placed in a thimble of a Soxhlet apparatus then extracted with ether for 48 h. Recrystallized from ether; orange powder; 92% yield; TLC R_f (EtOAc)= 0.45; mp 177–179 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.93, 0.95 (2t, *J*=7.4 Hz and *J*=7.5 Hz, 3H, CHCH₂CH₃), 1.46– 1.77 (m, 1H, CHCH₂CH₃), 2.09–2.34 (m, 1H, CHCH₂CH₃), 3.07 (s, 6H, N(CH₃)₂), 3.08–3.47 (m, 2H, CH₂CH), 3.52, 3.53 (2t, J=6.3 Hz and J=6.3 Hz, 3H, CHCH₂CH₃), 5.03, 5.05 (2dd, J=10.9, 2.1 Hz and J=10.7, 1.8 Hz, 1H, CH₂CH₂CH), 6.66, 6.78 (2t, J=1.4 Hz and J=1.3 Hz, 1H, CH); ¹³C NMR (CDCl₃, 50 MHz) δ 12.4 (CH₃), 23.0 (CH₂), 23.2 (CH₂), 42.8 (CH₃), 30.8 (CH₃), 47.7 (CH), 52.2 (CH₃), 52.5 (CH₃), 52.9 (CH₃), 59.6 (CH), 95.2, 95.5 (C), 109.7 (C), 144.5 (C), 145.6 (CH), 145.7 (C), 167.7 (C), 169.4 (C), 170.7 (C), 171.2 (C); IR: ν cm⁻¹ 1745, 1657, 1632, 1500, 1350. Anal. Calcd for C₂₀H₂₅O₉N₃: C, 53.21; H, 5.58; N, 9.31. Found: C, 53.20; H, 5.65; N, 9.27.

4.9. Dimethyl 7-(1-(methoxycarbonyl)propyl)-1-(acetoxymethylene)-1,2,3,5-tetrahydro-6-nitro-5-oxoindolizine-3,8-dicarboxylate (44)

Cerium trichloride (8 mg, 0.055 mmol) was added to a stirred solution of enamine 41 (50 mg, 0.11 mmol) in acetonitrile (0.1 mL), EtOAc (0.1 mL), and sodium metaperiodate in water (0.1 mL, 0.055 mmol). The mixture was stirred for 15 min then aqueous NaHCO₃ (1 mL) was added and the mixture was extracted by dichloromethane (2×10 mL). The combined organic layers were dried then evaporated. The residue was dissolved in a mixture of acetic anhydride (5 mL) and sodium acetate (3 mg, 0.022 mmol) and the solution was refluxed for 48 h. The residue obtained upon evaporation was diluted in water (3 mL) then was extracted with dichloromethane $(2 \times 10 \text{ mL})$. The combined organic layers were dried by MgSO₄ then evaporated; 96% yield; brown oil; TLC R_f (EtOAc)=0.7; ¹H NMR (CDCl₃, 200 MHz) δ 0.93, 0.96 (2t, J=7.4 Hz and J=7.5 Hz, 3H, CHCH₂CH₃), 1.51-1.77 (m, 1H, CHCH₂CH₃), 1.96-2.37 (m, 1H, CHCH₂CH₃), 2.27 (s, 3H, COCH₃), 3.04–3.39 (m, 2H, CH₂CH), 3.5 (t, J=7.0 Hz, 3H, CHCH₂CH₃), 3.69, 3.72 (2s, 3H, CO₂CH₃), 3.83, 3.84 (2s, 3H, CO₂CH₃), 3.87, 3.88 (2s, 3H, CO₂CH₃), 5.15 (dd, J=8.3, 1.4 Hz, 1H, CH₂CH₂CH), 7.95, 8.05 (2s, 1H, CH). This product was analyzed only by NMR.

References and notes

- (a) Smith, D. Comprehensive Organic Chemistry; Sammes, P. G., Ed.; Pergamon: Oxford, 1979; Vol. 4, p 3; (b) Bailey, T.; Goe, G.; Scriven, E. Heterocyclic Compounds; Newkome, G. R., Ed.; Wiley: New York, NY, 1984; Vol. 14, Part 5, p 1; (c) McKillop, A.; Boulton, A. Comprehensive Heterocyclic Chemistry; McKillop, A., Boulton, A., Eds.; Pergamon: Oxford, 1984; Vol. 2, p 67.
- (a) Pendrack, I.; Wittrock, R.; Kingbury, W. J. Org. Chem. 1995, 60, 2912; (b) Parreira, R. L. T.; Abrahao, O.; Galembeck, S. E. Tetrahedron 2001, 57, 3243; (c) Saari, W. S.; Hoffman, J. M.; Wai, J. S.; Fisher, T. E.; Rooney, C. S.; Smith, A. M.; Thomas, C. M.; Goldman, M. E.; O'Brien, J. A. J. Med. Chem. 1991, 34, 2922.
- Basf Aktiengesellschaft. PCT Int. Appl. WO 2002006233, 2002; Chem. Abstr. 2002, 136, 118386.
- Imperial Chemical Industries PLC. Eur. Pat. Appl. EP 259048, 1988; Chem. Abstr. 1988, 109, 73348.
- (a) Matsumoto, M.; Minato, H. *Tetrahedron Lett.* **1976**, *42*, 3827; (b) Cox, R.; O'Hagan, D. J. Chem. Soc., Perkin Trans. 1 **1991**, 2537; (c) Sumitomo Chemical Co. Eur. Pat. Appl. EP 535980, 1993; *Chem. Abstr.* **1993**, *119*, 117126; (d) Williams, D. R.; Lowder, P. D.; Gu, Y.-G. Tetrahedron Lett. **1997**, *38*, 327.

- 6. Brickner, S. Chem. Ind. 1997, 131.
- Seto, H.; Hayakawa, Y.; Shimazu, A. Jpn. Kokai Tokkyo Koho, JP 05306272, 1993.
- 8. Yuldashev, P. K. Chem. Nat. Compd. 2001, 37, 274.
- Kozikowski, A. P.; Campiani, G.; Sun, L.-Q.; Wang, S.; Saxena, A.; Doctor, B. P. J. Am. Chem. Soc. 1991, 118, 11357.
- (a) Wall, M. E.; Wani, M. C.; Cook, C. E.; Palmer, K. H.; McPhail, A. I.; Sim, G. A. J. Am. Chem. Soc. 1966, 88, 3888;
 (b) Schultz, A. G. Chem. Rev. 1973, 73, 365; Hutchinson, C. R. Tetrahedron 1981, 37, 1047; Chinese Drugs of Plant Origin; Tang, W., Eisenbrand, G., Eds.; Springer: New York, NY, 1992; p 239; Ciufolini, M. A.; Roschangar, F. Targets Heterocycl. Syst. 2000, 4, 25; Kim, D.-K.; Lee, N. Min. Rev. Med. Chem. 2002, 2, 611; Lerchen, H. G. Drugs Future 2002, 27, 869; Ulukan, H.; Swan, P. W. Drugs 2002, 62, 2039; Du, W. Tetrahedron 2003, 59, 8649; Lee, K. H. J. Nat. Prod. 2004, 67, 273; Craig, J. T.; Rahier, N. J.; Hecht, S. M. Bioorg. Med. Chem. 2004, 12, 1585; (c) Ciufolini, M. A.; Roschangar, F. Tetrahedron 1997, 53, 11049.
- 11. Schultz, A. Chem. Rev. 1973, 73, 1047.
- 12. Hutchinson, C. R. Tetrahedron 1981, 37, 1047.
- (a) Lavelle, F.; Bissery, M. C.; André, S.; Roquet, F.; Riou, J. F. Semin. Oncol. 1996, 23, 13; (b) Rothenberg, M. L.; Rowinsky, E. K.; Kuhn, J. G.; Burris, H. A.; Donehower, R. C.; Vonhoff, D. D. Camptothecins: New Anticancer Agents; Potmesil, M., Pinedo, H., Eds.; CRC: Boca Raton, FL, 1995; p 75; (c) Santos, A.; Zanetta, S.; Cresteil, T.; Debroussent, A.; Pein, F.; Raymont, E.; Vernillet, L.; Rise, M.; Bioige, V.; Gaugette, A.; Vassal, G. Clin. Cancer Res. 2000, 6, 2012.
- (a) Staker, B. L.; Feese, M. D.; Cushman, M.; Pommier, Y.; Zembower, D.; Stewart, L.; Burgin, A. B. *J. Med. Chem.* **2005**, 48, 2336; (b) Staker, B. L.; Hjerrild, K.; Feese, M. D.; Behnke, C. A.; Burgin, A. B.; Stewart, L. *Proc. Natl. Acad. Sci. U.S.A.* **2002**, *99*, 15387.
- (a) Sugimori, M.; Ejima, A.; Ohuki, S.; Uoto, K.; Mitsui, I.; Matsumoto, K.; Kawato, Y.; Yasuoka, M.; Sato, K.; Tagawa, H.; Terasawa, H. J. Med. Chem. 1994, 37, 3033; (b) Luzzio, M. J.; Besterman, J. M.; Emerson, D. L.; Evans, M. G.; Lackey, K.; Leitner, P. L.; McIntyre, G.; Morton, B.; Myers, P. L.; Peel, M.; Sisco, J. M.; Sternbach, D. D.; Tong, W.-Q.; Truesdale, A.; Uehling, D. E.; Vuong, A.; Yates, Y. J. Med. Chem. 1995, 38, 395; (c) Sugimori, M.; Ejima, A.; Ohsuki, S.; Uoto, K.; Mitsui, I.; Kawato, Y.; Hirota, Y.; Sato, K.; Terasawa, H. J. Med. Chem. 1998, 41, 2308; (d) Lavergne,

O.; Lesueur-Ginot, L.; Pla Rodas, F.; Bigg, D. C. H. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 2235; (e) Concerning this paradigm, see also: Hautefaye, P.; Cimetière, B.; Pierre, A.; Léonce, S.; Hickman, J.; Laine, W.; Bailly, C.; Lavielle, G. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 2731.

- Nicholas, A.; Wani, M.; Manikumar, G.; Wall, M.; Kohn, K.; Pommier, Y. J. Med. Chem. 1990, 33, 972.
- (a) Lavergne, O.; Lesueur-Ginot, L.; Pla Rodas, F.; Kasprzyk, P. G.; Pommier, J.; Demarquay, D.; Prevost, G.; Ulibarri, G.; Rolland, A.; Schiano-Liberatoir, A. M.; Harnett, J.; Pons, D.; Camara, J.; Biggs, D. C. H. J. Med. Chem. 1998, 41, 5410; (b) Lesueur-Ginot, L.; Demarquay, D.; Kiss, R.; Kasprzyk, P. G.; Dassonville, L.; Bailly, C.; Camara, J.; Lavergne, O.; Biggs, D. C. Cancer Res. 1999, 59, 2939; (c) Philipart, P.; Harper, L.; Chaboteaux, C. Clin. Cancer Res. 2000, 6, 1557; (d) Redinbo, M. R.; Steward, L.; Kuhn, P.; Champoux, J. J.; Hol, W. G. J. Science 1998, 279, 1504; (e) Pommier, Y. Nat. Rev. Cancer 2006, 6, 789.
- 18. Cagir, A. J. Am. Chem. Soc. 2003, 123, 13628.
- (a) Brunin, T.; Hénichart, J.-P.; Rigo, B. *Tetrahedron* 2005, 61, 7916;
 (b) Brunin, T.; Hénichart, J.-P.; Legentil, L.; Rigo, B. *Tetrahedron* 2006, 62, 3959;
 (c) Malecki, N.; Houssin, R.; Hénichart, J.-P.; Couturier, D.; Petra, F.; Legentil, L.; Rigo, B. J. Heterocycl. Chem. 2003, 40, 45.
- Shen, W.; Coburn, C.; Bornmann, W.; Danishefsky, S. J. Org. Chem. 1993, 58, 611.
- Katritzky, A.; Scriven, E.; Majumder, S.; Akhmedova, R.; Akhmedov, N.; Vakulenko, A. *Arkivoc* 2005, 179.
- 22. Schön, R.; Weiland, J. Steroids 1997, 62, 762.
- We have already described oxidation reaction encountered while heating compounds in aqueous hydrochloric or hydrobromic acid: Akué-Gédu, R.; Bourry, A.; Camus, F.; Norberg, B.; Durant, F.; Couturier, D.; DeBacker, M.; Rigo, B. *Heterocycles* 2004, *63*, 1855.
- Akué-Gédu, R.; Rigo, B. *Targets Heterocycl. Syst.*, in press; *Tetrahedron Lett.* **1989**, *30*, 7057; *Tetrahedron Lett.* **1996**, *37*, 485; Mashimov, V. I. *Tetrahedron* **1965**, *21*, 687; Rapoport, H. *Lect. Heterocycl. Chem.* **1978**, *IV*, S-47.
- (a) Dragovich, P.; Zhou, R.; Prins, T. J. Org. Chem. 2002, 67, 741; (b) Dragovich, P. J. Med. Chem. 2003, 46, 4572; (c) Zhang, X.; Schmitt, A.; Decicco, C. Tetrahedron Lett. 2002, 43, 9663; (d) Aberg, V.; Almqvist, F. Bioorg. Med. Chem. 2006, 14, 7563; (e) Millet, R.; Domarkas, J.; Rombaux, P.; Rigo, B.; Houssin, R.; Hénichart, J.-P. Tetrahedron Lett. 2002, 43, 5087.