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Tetrahedron

Toward new camptothecins. Part 4: On the reactivity of nitro and amino precursors of aza analogs of 5-methoxycarbonyl camptothecin

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> Received 1 June 2007; revised 21 June 2007; accepted 27 June 2007 Available online 4 July 2007

Abstract—In the context of formation of new aza analogs of camptothecin, nitration then reduction of condensed pyridones was realized, leading to new derivatives of pyrrolo-aza-indoles. Treatment of these compounds with hydrobromic acid led to new structure rearrangements while oxidation of the α -position was unsuccessful. Mechanisms of formation of the new products are discussed. $© 2007 Elsevier Ltd. All rights reserved.$

1. Introduction

The 2-pyridone unit is a key structural feature of a number of biologically active compounds.^{[1](#page-7-0)} Some pyridones act as antiviral, $\frac{3}{7}$ $\frac{3}{7}$ $\frac{3}{7}$ or as herbicides, $\frac{3}{7}$ pesticides, $\frac{4}{7}$ $\frac{4}{7}$ $\frac{4}{7}$ fungicides, $\frac{5}{7}$ $\frac{5}{7}$ $\frac{5}{7}$ antibio-tics,^{[6](#page-8-0)} or antioxidant agents.^{[7](#page-8-0)} On the other hand, it can be found in the toxin ricinine,^{[8](#page-8-0)} the psychoactive huperzine $A⁹$ $A⁹$ $A⁹$ or in the lead anticancer alkaloid camptothecin (Fig. 1).

The isolation and structure of camptothecin were reported in 1966 by Wall^{10a} and many reviews, devoted to this com-pound, revealed its paramount importance.^{[10b,c,12](#page-8-0)} Interest in this cytotoxic drug and semisynthetic analogs was stimulated when its mode of action was discovered. When the cleavable complex between topoisomerase I and DNA is stabilized by camptothecin, collision of the replication fork with this reversible complex^{[11](#page-8-0)} leads to cell death by prevent-ing DNA religation.^{[12](#page-8-0)} Irinotecan and topotecan have emerged from bioavaibility and tolerance studies, and are used in cancer treatment.^{[13](#page-8-0)} Recently, the crystal structure of the DNA–topoisomerase I–camptothecin complex was solved and two models of camptothecin–DNA–topoisomer-ase I interaction were formulated.^{[14](#page-8-0)} In the camptothecin series, structure–activity relationships are well established in regard to modifications in A, B, C, and D rings. They

Figure 1. Structures of some active pyridones.

also indicated the ability of substituents in positions 7, 9, 10, and 11 to maintain or improve biological activity. The need for a hydroxy lactone ring E has been established as a golden rule^{[15](#page-8-0)} and, for instance, lactams 1 [\(Fig. 2\)](#page-1-0) possess poor activity.[16](#page-8-0) However, this dogma fell with the discovery of other compounds stabilizing the complex between DNA and topoisomerase I in the same way as camptothecin: the homocamptothecins such as the diflomotecan, or the cyclopentanones such as $S39625$ are now in clinical phase.^{[17](#page-8-0)} The luotonin is also now known to exhibit one tenth of the camptothecin topo I inhibitory activity (Fig. 2).^{[18](#page-8-0)}

The targets of the present work were lactams 2 designed to explore the potential of aza analogs of cyclopentanones such as S39625 ([Fig. 2\)](#page-1-0). We have already described the synthesis of 5-methoxycarbonyl camptothecin 3 from DL meth-yl pyroglutamate 4.^{[19c](#page-8-0)} A key intermediate of this reaction scheme was ketone 5, obtained by oxidation of enamine 6 ([Scheme 1](#page-1-0)). Interest in the methoxycarbonyl group of 2 came from its position face to face with the major groove

Keywords: 2-Pyridone; Camptothecin; Decarboxylation; Nitro displacement.

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^{0040–4020/\$ -} see front matter © 2007 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2007.06.083

Figure 2. Camptothecin analogs.

in the potential complex with DNA and topo I. It can probably be replaced easily by a hydrogen, $19a$ thereby increasing the size of the library of compounds to be tested biologically.

Scheme 1. Synthesis of 5-methoxycarbonyl camptothecin analog.

2. Results and discussion

2.1. Synthesis of nitro compounds and treatment of products in acidic media

As a first approach to the reaction of nitration, pyridone 8^{19a} 8^{19a} 8^{19a} was submitted to the action of $AcONO₂$, although the preferential reactivity of these condensed pyridones toward electrophiles in the para position of the carbonyl was known.^{[20](#page-8-0)} Compound 9 nitrated at the *ortho* position was not observed in the reaction media, and only products 10– 12 were isolated by chromatography on $SiO₂$. The syntheses of mono and dinitro products 10 and 12 were not unexpected, but formation of the gem disubstituted compound 11 was interesting, highlighting the particular reactivity at the α -position of condensed pyridones (Scheme 2).

Because mono nitropyridone 9 was not formed during this first reaction, nitration of triester 7 was now realized. Only small amounts of nitrated triester 13 were isolated after using 63% HNO₃/AcOH or HNO₃/H₂SO₄; however, HNO₃/ TFAA^{[21](#page-8-0)} led to 79% of **13**, and $\overline{HNO_3/AC_2O^{22}}$ gave quantitative yield of the same product [\(Scheme 3\)](#page-2-0).

Removal of the aromatic methoxycarbonyl group of 13 was then studied. Such a decarboxylation of esters of condensed pyridones is well known, realized by reflux in 48% HBr. Thereby triester 13 was treated in these conditions ([Scheme](#page-2-0) [3\)](#page-2-0). The remaining ester functions were hydrolyzed in the course of the reaction and the compounds need to be reesterified; that was realized with MeOH, while drying the ternary azeotrope $H_2O/MeOH/CHCl₃$ on molecular sieves.^{[19a](#page-8-0)} Compounds 14–17 were obtained with a 95% total yield.

2.2. Formation of pyridone 14

Removal of the nitro group of the previous nitropyridone 13 is only a consequence of the reversibility of the reaction of electrophiles with aromatics. An interesting result is the better

Scheme 2. Reaction conditions: (i) (a) 48% HBr, reflux, 5 h; (b) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h;^{[19a](#page-8-0)} (ii) Ac₂O/63% HNO₃, 60 °C, 2 h.

Scheme 3. Reaction conditions: (i) $Ac_2O/63\%$ HNO₃, 60 °C, 1 h; (ii) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h.

yield of 14 obtained by using 30% HCl. This led very cleanly to 90% of 14, while 4% H₂SO₄ gave 70% of the same product, accompanied by low amounts of diester 15 and lactone 16. These better yields were not caused by the rise in the acidity because 30% HCl is less acidic than 48% HBr. So this increase in 14 was due to the decrease in the formation of the other by-products. This interpretation of the results was validated when KBr was added to HBr, leading to the preferential formation of lactone 16, to the disadvantage of 14 (Scheme 3).

2.3. Formation of oxidized products 15 and 16

Pyridones 15 and 16 came from an oxidation of the lateral chain of 13. In a possible mechanism that was due to oxygen

dissolved in hydrobromic acid^{[23](#page-8-0)} even when the reactions were performed under nitrogen. Because triester 7 (or the corresponding triacid) was not oxidized in the same conditions¹⁹ (Scheme 4), the nitro group of 13 was necessary for the oxidation to occur. As the presence of KBr increased the yield in lactone 16 (Scheme 3), the Br⁻ anion probably mediated this reaction. Alcohol 18 was thus obtained more easily from triester 13 or triacid 19 (Scheme 4). In this product, the hydroxyacid chain was substituted in the a-position of the acid by the strongly withdrawing nitropyridone group. Decarboxylation of acid 18, then cyclization of alcohol 20 led to lactone 16. On the other hand, alcohol 18 can evolve to ketone 21. Indeed, decarbonylation of α -amino (amido, oxo) acid derivatives 22 to give oxonium ions 23 is

Scheme 4. Reaction conditions: (i) 48% HBr, reflux, 5 h; (ii) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h.^{[19a](#page-8-0)}

a well-known reaction^{[24](#page-8-0)} (Scheme 5). When R=H (22), the oxonium formed was the protonated form of ketone 21. Another oxidation step changed the ethyl ketone chain of 21 to a new carboxylic acid 24; protolysis of the nitro group, decarboxylation of the para-carboxyacid, and reesterification led then to pyridone 15.

Scheme 5. Oxonium ions.

2.4. Formation of oxidized products 17

A first light on the reaction mechanism leading to compound 17 was obtained in the following reaction: in the hope to clarify the possible influence of ester groups, triester 13 was first heated in sodium hydroxide. The resulting crude product was heated at reflux in 48% HBr then reesterified in the usual way. Only new pyridone 25 was isolated in 30% yield, without traces of phenol 17. A possible mechanism involves initial nucleophilic attack of the hydroxyl ion on the carbon carrying the nitro function, with formation of the anionic σ^H adduct 26 (Meisenheimer complex). Upon reflux in HBr, the pyridone ring of triacid 27 would be open, to give ketone 28 (drawn in its enolic form in Scheme 6). Recyclization, oxidative aromatization, and decarboxylation would then furnish diacid 29, whose esterification gave 25.

In this process, the crucial need for the aromatic carboxylic group of 27 to open the pyridone ring was highlighted by the fact that analog 30, obtained from saponification of diester 17, does not evolve toward 25 (Scheme 6).

Formation of compound 17 can now be described as the opening of the pyridone ring of triacid 31 then hydrolysis of nitro compound 32 to give ketone 33 (these two compounds are drawn in their enolic forms in Scheme 6). Because 33, unlike 28, did not yield diester 25, the decarboxylation of 32 occurred before hydrolysis of the nitro group.

2.5. Reduction then decarboxylation of nitropyridone 13

Because decarboxylation of triester 13 does not yield the required nitropyridone, it was decided to perform this step after reduction of the nitro function. Treatment of 13 either with Pd/C or with Zn/HBr yields 83 or 70%, respectively, of azaindolinedione 34, thus furnishing the E ring of compound 2. Decarboxylation of 34 was realized as previously by reflux in 48% HBr then reesterification of acid 36 with MeOH. Ester 36 was not obtained, but methoxyazaindolinone 37 was isolated in 86% yield, accompanied by 2% of amine 38. Iminoether 39, formed from 36 and MeOH/H+, could be an intermediate in the formation of 37. On the other hand, oxidation of heterocycle 35 or of the aminoacid obtained from hydrolysis of the lactam ring of 35, probably yielded 40. As shown in [Scheme 7](#page-4-0), loss of carbon monoxide would then lead to aminoketones 41 then 38. Acid 35 was characterized by NMR, but was not isolated; if needed, it will be necessary to proceed to its esterification by using other methods that the reflux in MeOH/H⁺.

2.6. Attempts to introduce a ketone group in C ring

Many reactions were performed in the aim to introduce a ketone functionality in the pyrrolidine C ring. Some of them are reported in [Scheme 8;](#page-4-0) the use of selenium oxide was earlier reported by Danishefsky²⁰ in his synthesis of camptothecin, and we have already described the oxidation of enedimethylamines in the same context[.19](#page-8-0) Only one result needs some comments: heating nitropyridone 13 with Bredereck's reagents easily gave 92% of enamine 42. Oxidation of this compound by NaIO_4 led to entire degradation of the reaction medium. Because that could be caused by oxidative cleavage of the enol form of ketone 43, stabilization of the keto form was attempted by complexation with $CeCl₃$. The obtained mixture led rapidly to a very complex NMR spectrum (there were three asymmetric centers in the new product obtained) but some peaks could originate from an aldehyde or its enol form. The crude mixture was then

Scheme 6. Reaction conditions: (i) 48% HBr, reflux, 5 h; (ii) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 \AA , reflux, 48 h; (iii) NaOH, reflux 6 h.

Scheme 7. (i) Zn/HBr/MeOH, 20 °C; 1 h; (ii) 48% HBr, reflux, 24 h; (iii) MeOH, MeSO₃H, CHCl₃, molecular sieves 3 Å, reflux, 48 h.

Scheme 8. α-Oxidation of C ring.

treated by acetic anhydride, giving a quantitative yield of enol acetate 44.

2.7. NMR spectra of new compounds

Many of the new products obtained in this study were unexpected compounds. Elucidation of their structures was

Figure 3. ¹H NMR spectra of pyridones 11, 15–17, 29, and 39 and assignments determined by 2D NMR techniques: COSY, NOESY, HMBC, and HSQC.

carried out by NMR, mass spectrometry (LC–MS), infra-red and elemental analyses. Some of ¹H NMR spectra as well as HMBC and HSQC correlations are reported in Figure 3.

3. Conclusion

Introduction of the ketone functionality in the pyrrolidine C ring of the key precursors of aza analogs of 5-methoxycarbonyl camptothecin was unsuccessful. Thus it will be necessary to use other methods than nitration to obtain these compounds. However, some of the new products described in this study could give dipeptide analog scaffolds useful in the field of rigid β -turn peptidomimetics [\(Fig. 4\)](#page-5-0).^{[25](#page-8-0)}

Figure 4. Analogy between 34, 38, and 39 and rigid β -turn peptidomimetics.

4. Experimental section

4.1. General

Melting points were determined using an Electrothermal apparatus and are uncorrected. ¹H and ¹³C NMR spectra were obtained on a Varian Gemini 2000 at 200 and 50 MHz, respectively. IR spectra were obtained in ATR mode on a FTIR Bruker Tensor 27. Thin layer chromatographies were performed on precoated Kieselgel $60F_{254}$ plates. APCI⁺ (atmospheric pressure chemical ionization) mass spectra were obtained on an LC–MS system Thermo Electron Surveyor MSQ. Microanalyses were performed by the 'Service de Microanalyses' of LSEO, Université de Bourgogne, Dijon, France. All products are obtained as mixtures of diastereoisomers.

4.2. Nitration of pyridone 8

Nitric acid (63%, 2 mL) was slowly added to a solution of pyridone 8 (900 mg, 3 mmol) in acetic anhydride (11 mL) at 0 °C. The mixture was stirred at 60 °C for 1 h and the solvent was evaporated. The residue was diluted with EtOAc (50 mL) and washed with satd aqueous NaHCO_3 (50 mL). The aqueous phase was extracted with EtOAc $(3\times30 \text{ mL})$ and the combined organic layers were dried by MgSO₄ then evaporated. The residue was purified by chromatography on $SiO₂$ (heptane/EtOAc, $100/0\%$ to $0/100\%$, to give compounds $10-12$.

4.2.1. Methyl 7-(1-(methoxycarbonyl)propyl)-1,2,3,5-tetrahydro-6,8-dinitro-5-oxoindolizine-3-carboxylate (10). Recrystallized from methanol/ether; yellow powder; 40% yield; TLC R_f (EtOAc)=0.7; mp 65–67 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.92, 0.93 (2t, J=7.5 and 7.5 Hz, 3H, CHCH₂CH₃), 1.52–1.78 (m, 2H, CHCH₂CH₃), 2.20–2.69 $(m, 2H, CH_2CH_2CH), 3.44-3.78$ $(m, 3H, CH_2CH_2CH,$ $CHCH_2CH_3$), 3.65 (s, 3H, CO₂CH₃), 5.19, 5.22 (2dd, J=9.6, 3.3 Hz and J=8.0, 1.6 Hz, 1H, CH₂CH₂CH); ¹³C NMR (CDCl₃, 50 MHz) δ 12.2 (CH₃), 25.0 (CH₂), 25.4 (CH₂), 33.4 (CH₂), 48.7 (CH), 52.2 (CH₃), 53.0 (CH₃), 62.3 (CH), 118.2 (C), 146.0 (C), 153.3 (C), 159.2 (C), 169.3 (C), 171.3 (C), 171.6 (C); IR: ν cm⁻¹ 1745, 1680, 1534, 1338. Anal. Calcd for C₁₅H₁₇O₉N₃: C, 47.00; H, 4.47; N, 10.96. Found: C, 46.92; H, 4.66; N, 10.63.

4.2.2. Methyl 7-(1-(methoxycarbonyl)propyl)-1,2,3,5-tetrahydro-1,1,8-trinitro-5-oxoindolizine-3-carboxylate (11). Recrystallized from EtOAc/ether; yellow powder; 10% yield; mp 105–107 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.81, 0.83 (2t, J=7.3 Hz and J=7.3 Hz, 3H, CH₂CH₃), 1.71–2.20 $(m, 2H, CH_3CH_2CH), 3.65, 3.67 (2s, 3H, CO_2CH_3), 3.68$ $(t, J=7.3 \text{ Hz}, 1H, CHCH_2CH_3), 3.75, 3.76 (2s, 3H, CO_2CH_3),$ 3.78–4.14 (m, 2H, CH₂CH), 5.43, 5.45 (2dd, $J=9.5$, 1.7 Hz and $J=9.4$, 1.6 Hz, 1H, CH₂CH₂CH), 7.3, 7.31 (2s, 1H, CH); ¹³C NMR (CDCl₃, 50 MHz) δ 11.6 (CH₃), 24.4 (CH_2) , 24.8 (CH₂), 35.5 (CH), 47.9 (CH₃), 53.0 (CH₃), 106.4 (C), 119.4 (C), 139.6 (CH), 139.7 (C), 144.9 (C), 151.8 (C), 167.7 (C), 170.4 (C); IR: ν cm⁻¹ 1741, 1683, 1632, 1543, 1358; LC-MS (APCI⁺) m/z 429 (MH⁺). Anal. Calcd for $C_{15}H_{16}N_4O_{11}$: C, 42.06; H, 3.77; N, 13.08. Found: C, 42.38; H, 3.80; N, 13.48.

4.2.3. Methyl 7-(1-(methoxycarbonyl)propyl)-1,2,3,5-tetrahydro-8-nitro-5-oxoindolizine-3-carboxylate (12). Recrystallized from methanol/ether; yellow powder; 35% yield; TLC R_f (EtOAc)=0.8; ¹H NMR (CDCl₃, 200 MHz) δ 0.99, 1 (2t, $J=7.5$ Hz and $J=7.5$ Hz, 3H, CH₂CH₃), 1.75– 2.27 (m, 1H, CH₂CH₂CH), 2.3–2.7 (m, 1H, CH₂CH₂CH), 3.55–3.67 (m, 2H, CH_2CH_2CH), 3.71 (s, 3H, CO_2CH_3), 3.83 (s, 3H, CO_2CH_3), 3.92, 3.94 (2t, J=7.1 Hz and $J=7.2$ Hz, 1H, CHCH₂CH₃), 5.18 (dd, $J=9.3$, 3.4 Hz, 1H, CH_2CH_2CH , 6.44 (s, 1H, CH); ¹³C NMR (CDCl₃, 50 MHz) δ 11.8 (CH₃), 24.9 (CH₂), 27.3 (CH₂), 29.4 $(CH₂), 45.1$ (CH), 52.0 (CH₃), 52.8 (CH₃), 61.2 (CH), 100.6 (CH), 126.7 (C), 138.3 (C), 141.7 (C), 156.5 (C), 170.1 (C), 173.6 (C). Anal. Calcd for $C_{15}H_{18}O_7N_2$: C, 53.25, H, 5.36, N, 8.28. Found: C, 53.11; H, 5.42; N, 8.08.

4.3. Dimethyl 7-[1-(methoxycarbonyl)propyl]-6-nitro-5 oxo-1,2,3,5-tetrahydro-3,8-indolizinedicarboxylate (13)

Nitric acid (63%, 90 mL, 0.6 mol) was slowly added to a solution of compound 7 (70 g, 0.2 mol) in acetic anhydride (1 L) at 0 °C. The mixture was stirred at 60 °C for 1 h and the solvent was evaporated. The residue was diluted with EtOAc (50 mL) and washed by satd aqueous NaHCO₃ (50 mL). The aqueous phase was extracted with EtOAc $(3\times30 \text{ mL})$ and the combined organic layers were dried (MgSO4). The residue obtained upon evaporation was crystallized from methanol/ ether; yellow powder; 95% yield; TLC R_f (EtOAc)=0.6; mp 147–149 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.92, 0.95 $(2t, J=7.3 \text{ Hz and } J=7.6 \text{ Hz}, 3H, CH_2CH_3), 1.5-1.7 \text{ (m)}$ 2H, CHCH₂CH₃), 2.25–2.65 (m, 2H, CH₂CHCO), 3.35– 3.65 (m, 3H, CHCH₂CH₃, CH₂CH₂CH), 3.68, 3.69 (2s, 3H, $CO₂CH₃$), 3.79, 3.80 (2s, 3H, $CO₂CH₃$), 3.83, 3.84 (2s, 3H, $CO₂CH₃$), 5.2, 5.24 (2dd, J=9.6, 3.5 Hz and J=9.6, 2.9 Hz, 1H, NCHCO₂CH₃); ¹³C NMR (CDCl₃, 50 MHz) δ 12.6 (CH_3) , 23.7 (CH₂), 24.1 (CH₂), 25.5 (CH₂), 33.3 (CH₂), 48.1 (CH), 52.1 (CH3), 52.4 (CH3), 53.3 (CH3), 62.6 (CH), 104.6 (C), 144.0 (C), 152.7 (C), 157.3 (C), 164.6 (C), 169.1 (C), 170.4 (C); IR: ν cm⁻¹ 1740, 1723, 1660, 1605, 1533, 1433. Anal. Calcd for $C_{17}H_{20}O_9N_2$: C, 51.51; H, 5.02; N, 6.79. Found: C, 51.52; H, 5.09; N, 7.07.

4.4. Decarboxylation of nitro compound 13

A stirred solution of pyridone 7 (1 g, 2.5 mmol) in 48% hydrobromic acid (50 mL) was heated at 130 °C for 48 h then evaporated. Methanesulfonic acid (0.10 mg) in methanol (200 mL) and chloroform (200 mL) were added to the

residue, and the mixture was refluxed for 48 h while drying the solvent by condensing it in a Soxhlet-type apparatus containing 3 Å molecular sieves (50 g) . Dichloromethane (200 mL) was added to the residue obtained upon evaporation, and the solution was washed with aqueous NaHCO₃. The organic phase was dried $(MgSO₄)$ then evaporated. The residue was purified by chromatography on $SiO₂$ (heptane/EtOAc, 100/0% to 0/100%) to give pyridones 14–17.

4.4.1. Methyl 7-[1-(methoxycarbonyl)propyl]-5-oxo-1,2,3,5-tetrahydroindolizine-3-carboxylate (14). Recrystallized from EtOAc; white powder; 48% yield; TLC R_f $(EtOAc)=0.45$; mp 65–67 °C; ¹H NMR $(CDCl_3, 200 MHz)$ δ 0.91, 0.92 (2t, J=7.2 Hz and J=7.6 Hz, 3H, CHCH₂CH₃), 1.7–2.1 (m, 2H, CHC H_2 CH₃), 2.2–2.6 (m, 2H, CH₂CH₂CH), 2.9–3.2 (m, 2H, CH₂CH₂CH), 3.28 (t, J=7.7 Hz, 1H, $CHCH_2CH_3$), 3.69, 3.69 (2s, 3H, CO₂CH₃), 3.8 (s, 3H, CO₂CH₃), 5.08, 5.09 (2dd, J=9.1, 2.9 Hz and J=5.9, 3.2 Hz, 1H, CH₂CH₂CH₁, 6.15 (s, 1H, ArH), 6.3 (s, 1H, ArH); ¹³C NMR (CDCl₃, 50 MHz) δ 11.8 (CH3), 25.5 (CH₂), 26.1 (CH2), 30.3 (CH2), 52.1 (CH), 52.6 (CH3), 52.9 (CH3), 60.9 (CH), 101 (CH), 116.4 (CH), 150 (C), 152.6 (C), 161.3 (C) , 170.4 (C) , 172.8 (C) ; IR: ν cm⁻¹ 1727, 1657, 1589, 1532, 1432, 1204; Anal. Calcd for C₁₅H₁₉NO₅: C, 61.49; H, 5.91; N, 5.17. Found: C, 61.49; H, 6.78; N, 5.17.

4.4.2. Dimethyl 1,2,3,5-tetrahydro-5-oxoindolizine-3,7 dicarboxylate (15). White powder; 23% yield; TLC R_f $(EtOAC) = 0.4$; ¹H NMR (CDCl₃, 200 MHz) δ 2.24–2.74 (m, 2H, CH₂CH₂CH), 3.03-3.36 (m, 2H, CH₂CH₂CH), 3.79 (s, 3H, CO₂CH₃), 3.9 (s, 3H, CO₂CH₃), 5.13 (dd, J=9.4, 3.1 Hz, 1H, CH₂CH₂CH), 6.63, 7.03 (2s, ArH, 2H); ¹³C NMR (CDCl₃, 50 MHz) δ 8.4 (CH₃), 25.8 (CH₂), 27.4 (CH_2) , 29.5 (CH_2) , 52.9 (CH_3) , 61.6 (CH) , 81.2 (CH) , 102.3 (C), 108.7 (CH), 155.1 (C), 159.3 (C), 161.1 (C), 167.4 (C), 169.6 (C); LC-MS (APCI⁺) m/z 251 (MH⁺). Whatever the purification method, this product was always contaminated with 10% of 14.

4.4.3. Methyl 3-ethyl-1,3,5,7,8,9-hexahydro-1,5-dioxofuro[3,4-g]indolizine-7-carboxylate (16). Recrystallized from EtOAc/ether; white powder; 23% yield; TLC R_f $(EtOAc)=0.4$; mp 112–114 °C; ¹H NMR $(CDCl_3, 200$ MHz) δ 1.03 (t, J=7.4 Hz, 3H, CHCH₂CH₃), 1.69–2.16 (m, 2H, $CHCH_2CH_3$), 2.36–2.75 (m, 2H, CH₂CH₂CH), 3.34–3.69 $(m, 2H, CH_2CH_2CH), 3.82$ (s, 3H, CO_2CH_3), 5.17 (dd, $J=9.6$, 3.2 Hz, 1H, CH₂CH₂CH₁, 5.31, 5.34 (2dd, $J=7.0$) 1.4 Hz and $J=7.0$, 1.4 Hz, 1H, CHCH₂CH₃), 6.29 (m, 1H, ArH); ¹³C NMR (CDCl₃, 50 MHz) δ 8.4 (CH₃), 25.8 (CH₂), 27.4 (CH₂), 29.5 (CH₂), 52.9 (CH₃), 61.6 (CH), 81.2 (CH), 102.3 (C), 108.7 (CH), 155.1 (C), 159.3 (C), 161.1 (C), 167.4 (C), 169.6 (C); IR: ν cm⁻¹ 1742, 1677; LC-MS (APCI⁺) m/z 278 (MH⁺). Anal. Calcd for C₁₄H₁₅O₅N: C, 60.65; H, 5.45; N, 5.05. Found C, 60.60; H, 5.60; N, 5.08.

4.4.4. Methyl 7-(1-(methoxycarbonyl)propyl)-1,2,3,5-tetrahydro-6-hydroxy-5-oxoindolizine-3-carboxylate (17). Recrystallized from EtOAc; white cotton; 1% yield; TLC R_f (EtOAc)=0.55; mp 172–174 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.91, 0.93 (2t, J=7.4 Hz and J=7.4 Hz, 3H, CHCH₂CH₃), 1.59-1.85 (m, 1H, CHCH₂CH₃), 1.9-2.15 (m, 1H, CHC H_2CH_3), 2.23–2.65 (m, 2H, CH₂CH₂CH), 3.68, 3.7 (2s, CO₂CH₃, 3H), 3.8, 3.81 (2s, CO₂CH₃, 3H), 3.93, 3.95 (2t, $J=7.7$ Hz and $J=7.7$ Hz, 1H, CHCH₂CH₃), 5.11, 5.13 (2dd, $J=7.4$, 1.8 Hz and $J=7.4$, 1.8 Hz, 1H, CH_2CH_2CH , 6.16 (m, 1H, ArH), 6.96 (br s, 1H, OH); ¹³C NMR (CDCl₃, 50 MHz) δ 11.8 (CH₃), 24.9 (CH₂), 27.3 (CH_2) , 29.4 (CH₂), 45.1 (CH), 52.0 (CH₃), 52.8 (CH₃), CH (61.2), 100.6 (CH), 126.7 (C), 138.3 (C), 141.7 (C), 156.5 (C), 170.1 (C), 173.6 (C); IR: ν cm⁻¹ 1751, 1734, 1677, 1340, 1211; LC-MS (APCI⁺) m/z 310 (MH⁺). Anal. Calcd for $C_{15}H_{19}O_6N$: C, 8.25; H, 6.19; N, 4.53. Found: C, 58.25; H, 6.24; N, 4.74.

4.5. Dimethyl 6-ethyl-5-oxo-1,2,3,5-tetrahydroindolizine-3,7-dicarboxylate (29)

A stirred solution of pyridone 7 (500 mg, 1, 3 mmol) in 30% sodium hydroxide (20 mL) was heated at 70 °C for 16 h. The mixture was acidified by 48% hydrobromic acid (50 mL) and was heated at 130 $^{\circ}$ C for 48 h then evaporated. Methanesulfonic acid (0.10 mg) in methanol (200 mL) and chloroform (200 mL) were added to the residue, and the mixture was refluxed for 48 h while drying the solvent by condensing it in a Soxhlet-type apparatus containing 3 Å molecular sieves (50 g). Dichloromethane (200 mL) was added to the residue obtained upon evaporation, and the solution was washed with aqueous $NaHCO₃$. The organic phase was dried $(MgSO₄)$ then evaporated. The residue obtained upon evaporation was crystallized from methanol/ ether; yellow powder; 45% yield; ¹H NMR (CDCl₃, 200 MHz) δ 1.14 (t, J=7.4 Hz, 3H, CH₂CH₃), 2.22–2.61 $(m, 2H, CHCH₂CH₂), 2.97-3.30$ $(m, 2H, CH₂CH₂CH),$ 3.80 (s, 3H, CO₂CH₃), 3.90 (s, 3H, CO₂CH₃), 5.11 (dd, $J=9.3$ Hz and $J=3.5$ Hz, 1H, $CH_2CH_2CH_3$; ¹³C NMR (CDCl₃, 50 MHz) δ 13.4 (CH₃), 21.7 (CH₂), 26.3 (CH₂), 30.3 (CH2), 52.4 (CH3), 52.7 (CH3), 61.6 (CH), 99.9 (CH), 133.9 (C), 138.9 (C), 146.9 (C), 161.4 (C), 167.3 (C), 170.4 (C); LC-MS (APCI⁺) m/z 280 (MH⁺). Anal. Calcd for $C_{14}H_{17}O_5N$: C, 60.21; H, 6.14; N, 5.02. Found: C, 59.67; H, 6.21; N, 4.89.

4.6. Methyl 3-ethyl-2,9-dioxo-2,3,5,6,7,9-hexahydro-1Hpyrrolo[2,3-f]indolizine-4,7-dicarboxylate (34)

Zinc dust (600 mg) was slowly added to a stirring solution of pyridone 7 (400 mg, 1 mmol) in HBr (48%, 25 mL) and methanol (2 mL). After 1 h, the mixture was filtered on Celite and the filtrate was diluted with satd aqueous $NaHCO₃$ (50 mL). The aqueous layer was extracted by EtOAc $(3\times100 \text{ mL})$ and the combined organic layers were dried. Residue obtained upon evaporation was purified by crystallization in methanol. Recrystallized from methanol; white powder; 70% yields; TLC R_f (EtOAc)=0.56; mp 174– 176 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.83 (t, J=7.3 Hz, 3H, CH₂CH₃), 1.93–2 (m, 2H, CHCH₂CH₃), 2.3–2.66 (m, 2H, CH₂CHCO), 3.48–3.66 (m, 2H, CH₂CH₂CH), 3.83 (s, 3H, CO_2CH_3), 3.85–3.88 (m, 4H, CO_2CH_3 and $CHCH_2CH_3$), 5.27 (dd, J=3.3, 9.5 Hz, 1H, NCHCO₂CH₃), 8.6 (s, 1H, NH); ¹³C NMR (CDCl₃, 50 MHz) δ 9.2 (CH₃), 23.4 (CH₂), 26.0 (CH₂), 32.5 (CH₂), 50.7 (CH), 51.7 (CH₃), 53.0 (CH₃), 61.6 (CH), 104.4 (C), 130.5 (C), 135.6 (C), 151.2 (C), 151.4 (C), 164.7 (C), 169.9 (C), 178.6 (C); IR: ν cm⁻¹ 3224, 1729, 1717, 1653, 1576, 1537, 1438, 1212. Anal. Calcd for $C_{15}H_{17}O_9N_3$: C, 47.00, H, 4.47, N, 10.96. Found: C, 46.92; H, 4.66; N, 10.63.

4.7. Decarboxylation of compound 34

A stirred solution of pyridone 34 (6 g, 18 mmol) in 48% hydrobromic acid (120 mL) was heated at 130 $^{\circ}$ C for 24 h then evaporated. Methanesulfonic acid (0.25 mg) in methanol (200 mL) and chloroform (200 mL) were added to the residue and the solution was refluxed for 48 h while drying the solvent by condensing it in a Soxhlet-type apparatus containing $3 A$ molecular sieves (50 g). Dilute NaHCO₃ (200 mL) was added to the residue obtained upon evaporation and the solution was extracted with dichloromethane $(2\times400 \text{ mL})$. The combined organic layers were dried $(MgSO₄)$ then evaporated. The residue was purified by chromatography on $SiO₂$ (heptane/EtOAc, 100/0% to 0/100%) to give pyridones 37 and 38.

4.7.1. Methyl 3-ethyl-5,6,7,9-tetrahydro-2-methoxy-9 oxo-1H-pyrrolo[2,3-f]indolizine-7-carboxylate (37). Recrystallized from methanol/ether; gray powder; 97% yield; TLC R_f (EtOAc)=0.15; mp 235–237 °C; ¹H NMR (CDCl₃, 200 MHz) δ 1.16 (t, J=7.5 Hz, 3H, CH₂CH₃), 2.22–2.62 (m, 2H CH₂CH₂CH), 2.52 (q, J=7.5 Hz, 2H, CH₂CH₃), 2.97–3.32 (m, 2H, CH_2CH_2CH), 3.74 (s, 3H, CO_2CH_3), 4.08 (s, 3H, OCH₃), 5.17 (dd, J=9.4, 2.6 Hz, 1H, CH_2CH_2CH , 6.37 (t, J=1.4 Hz, 1H, ArH); ¹³C NMR (CDCl₃, 50 MHz) δ 15 (CH₂), 15.8 (CH₃), 27.4 (CH₂), 29.6 (CH_2) , 52.5 (CH₃), 59.3 (CH₃), 60.5 (CH), 95.7 (CH), 99.4 (C), 114.2 (C), 133.5 (C), 139.7 (C), 152.0 (C), 152.5 (C), 171.5 (C); IR: ν cm⁻¹ 1750, 1663, 1587, 1567, 1198; LC-MS (APCI⁺) m/z 291 (MH⁺). Anal. Calcd for C₁₅H₁₈O₄-N₂ H₂O: C, 58.43; H, 6.54; N, 9.09. Found: C, 58.25; H, 6.24; N, 9.60.

4.7.2. Methyl 6-amino-1,2,3,5-tetrahydro-5-oxo-7-propionylindolizine-3-carboxylate (38). Yellow foam; 2% yield; TLC R_f (EtOAc)=0.6; ¹H NMR (CDCl₃, 200 MHz) δ 1.19 (t, J=1.4 Hz, 3H, CH₂CH₃), 2.22–2.61 (m, 2H CH_2CH_2CH), 2.52 (q, J=7.4 Hz, 2H, CH_2CH_3), 2.90–3.22 (m, 2H, CH₂CH₂CH), 3.79 (s, 3H, CO₂CH₃), 5.11 (dd, J= 9.0, 3.1 Hz, 1H, CH₂CH₂CH₁, 6.4 (t, J=1.6 Hz, 1H, ArH), 7.24 (br s, 2H, NH₂); ¹³C NMR (CDCl₃, 50 MHz) δ 8.1 $(CH₃), 27.0 (CH₂), 28.8 (CH₂), 32.6 (CH₂), 52.6 (CH₃), 60.9$ (CH), 99.4 (CH), 112.7 (C), 131.3 (C), 140.4 (C), 157.2 (C), 170.3 (C), 202.0 (C). Anal. Calcd for $C_{14}H_{17}O_4N$: C, 59.08; H, 6.10; N, 10.60. Found: C, 59.64; H, 6.42; N, 10.02.

4.8. Dimethyl $(1E)$ -7- $(1$ -(methoxycarbonyl)propyl)-1-((dimethylamino)methylene)-1,2,3,5-tetrahydro-6 nitro-5-oxoindolizine-3,8-dicarboxylate (42)

A stirred mixture of triester 13 (300 mg, 0.75 mmol) and Bredereck's reagent (210 mg, 1.05 mmol) was heated at 110 °C for 3 h (\bar{N}_2). The residue obtained upon evaporation was diluted in water (40 mL) then extracted by dichloromethane $(2\times100 \text{ mL})$. The residue was dissolved in dichloromethane, Celite was added, and the solvent was evaporated. The powder was placed in a thimble of a Soxhlet apparatus then extracted with ether for 48 h. Recrystallized from ether; orange powder; 92% yield; TLC R_f (EtOAc)= 0.45; mp 177–179 °C; ¹H NMR (CDCl₃, 200 MHz) δ 0.93, 0.95 (2t, J=7.4 Hz and J=7.5 Hz, 3H, CHCH₂CH₃), 1.46– 1.77 (m, 1H, CHCH₂CH₃), 2.09–2.34 (m, 1H, CHCH₂CH₃), 3.07 (s, 6H, N(CH3)2), 3.08–3.47 (m, 2H, CH2CH), 3.52,

3.53 (2t, J=6.3 Hz and J=6.3 Hz, 3H, CHCH₂CH₃), 5.03, 5.05 (2dd, $J=10.9$, 2.1 Hz and $J=10.7$, 1.8 Hz, 1H, CH₂CH₂CH), 6.66, 6.78 (2t, $J=1.4$ Hz and $J=1.3$ Hz, 1H, CH); ¹³C NMR (CDCl₃, 50 MHz) δ 12.4 (CH₃), 23.0 (CH_2) , 23.2 (CH₂), 42.8 (CH₃), 30.8 (CH₃), 47.7 (CH), 52.2 (CH₃), 52.5 (CH₃), 52.9 (CH₃), 59.6 (CH), 95.2, 95.5 (C), 109.7 (C), 144.5 (C), 145.6 (CH), 145.7 (C), 167.7 (C), 169.4 (C), 170.7 (C), 171.2 (C); IR: ν cm⁻¹ 1745, 1657, 1632, 1500, 1350. Anal. Calcd for $C_{20}H_{25}O_9N_3$: C, 53.21; H, 5.58; N, 9.31. Found: C, 53.20; H, 5.65; N, 9.27.

4.9. Dimethyl 7-(1-(methoxycarbonyl)propyl)-1-(acetoxymethylene)-1,2,3,5-tetrahydro-6-nitro-5-oxoindolizine-3,8-dicarboxylate (44)

Cerium trichloride (8 mg, 0.055 mmol) was added to a stirred solution of enamine 41 (50 mg, 0.11 mmol) in acetonitrile (0.1 mL), EtOAc (0.1 mL), and sodium metaperiodate in water (0.1 mL, 0.055 mmol). The mixture was stirred for 15 min then aqueous NaHCO₃ (1 mL) was added and the mixture was extracted by dichloromethane $(2\times10$ mL). The combined organic layers were dried then evaporated. The residue was dissolved in a mixture of acetic anhydride (5 mL) and sodium acetate (3 mg, 0.022 mmol) and the solution was refluxed for 48 h. The residue obtained upon evaporation was diluted in water (3 mL) then was extracted with dichloromethane $(2\times10 \text{ mL})$. The combined organic layers were dried by MgSO₄ then evaporated; 96% yield; brown oil; TLC R_f (EtOAc)=0.7; ¹H NMR (CDCl₃, 200 MHz) δ 0.93, 0.96 (2t, J=7.4 Hz and J=7.5 Hz, 3H, $CHCH_2CH_3$, 1.51–1.77 (m, 1H, CHCH₂CH₃), 1.96–2.37 (m, 1H, CHCH₂CH₃), 2.27 (s, 3H, COCH₃), 3.04–3.39 (m, 2H, CH₂CH), 3.5 (t, J=7.0 Hz, 3H, CHCH₂CH₃), 3.69, 3.72 (2s, 3H, CO_2CH_3), 3.83, 3.84 (2s, 3H, CO_2CH_3), 3.87, 3.88 (2s, 3H, CO₂CH₃), 5.15 (dd, J=8.3, 1.4 Hz, 1H, CH_2CH_2CH , 7.95, 8.05 (2s, 1H, CH). This product was analyzed only by NMR.

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